

Developing a CRISPR-based Prime Editing Workflow for Human Induced Pluripotent Stem Cells to Model Cardiac Arrhythmogenic *TNNT2* Variants

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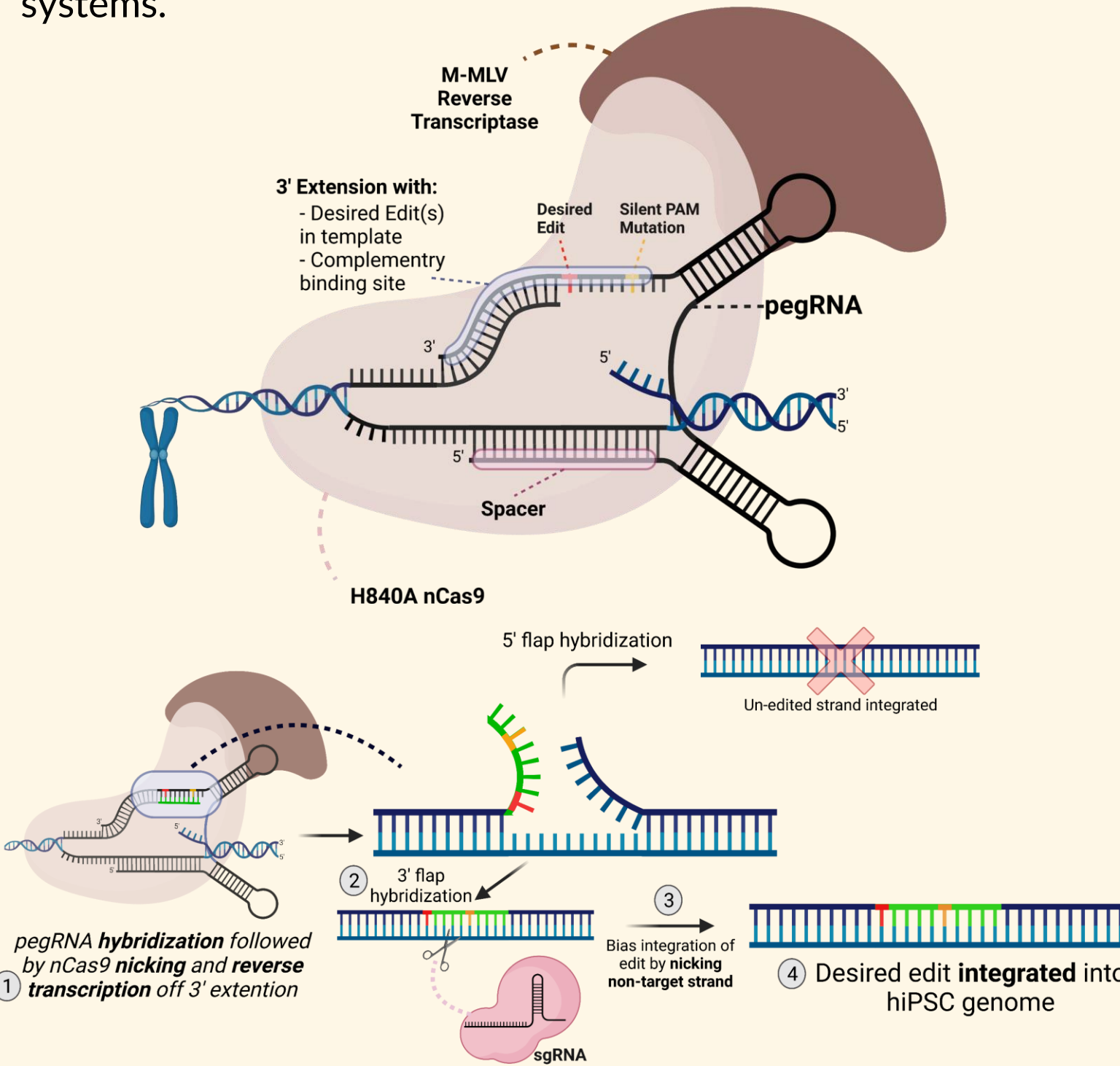
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Background

Hypertrophic cardiomyopathy (HCM) is the most prevalent inherited cardiovascular disease.

- Affect 1:500 individuals and the primary cause of sudden cardiac arrest (SCA) in youth and young adults, including elite athletes.
- Interestingly, some patient specific *TNNT2* variants related HCM develop varying degrees of HCM yet demonstrate a disproportionately high incidence of SCA.

CRISPR-based prime editing builds on established genome editing protocols with the benefits of using a nicking Cas9 endonuclease circumventing the array of issues arising from the double-strand breaks (DSBs) of traditional CRISPR/Cas9 systems.



3 components of PE3b system:

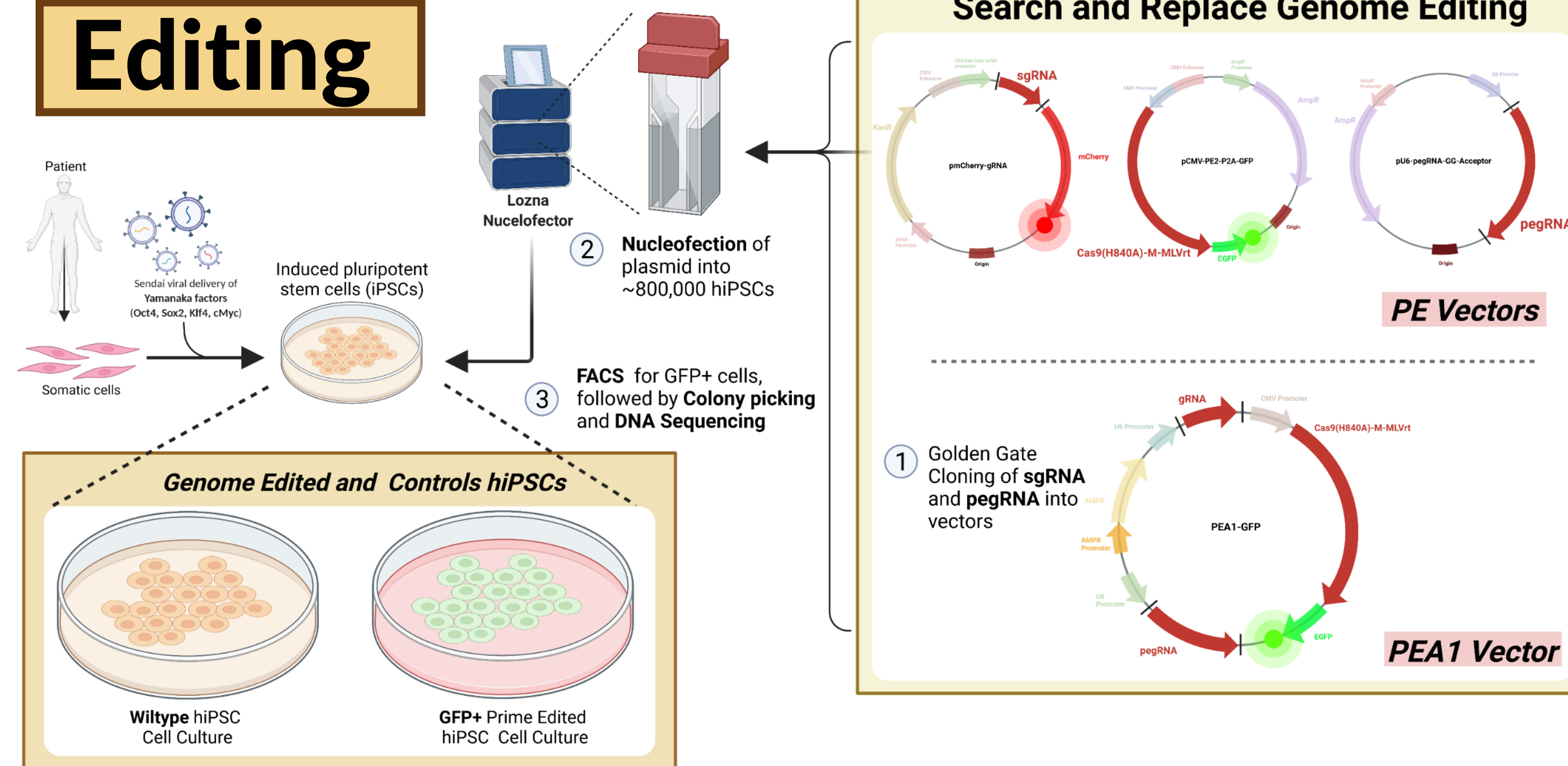
1. Cas9 endonuclease (H840A Cas9n) fused to a Moloney murine leukemia modified reverse transcriptase (M-MLV-RT)
2. Prime editing guide RNA (pegRNA)
3. Nicking guide RNA (ngRNA)

Research Objective

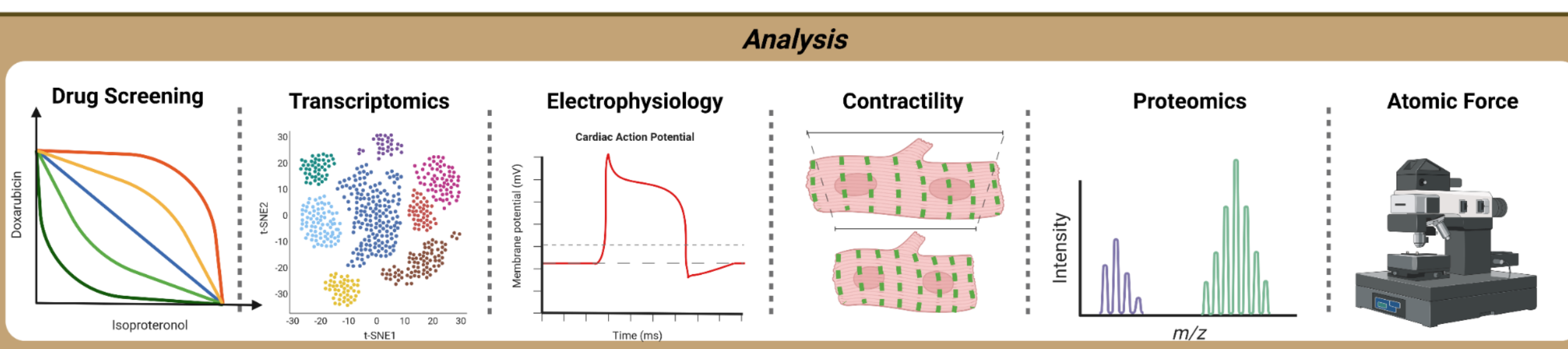
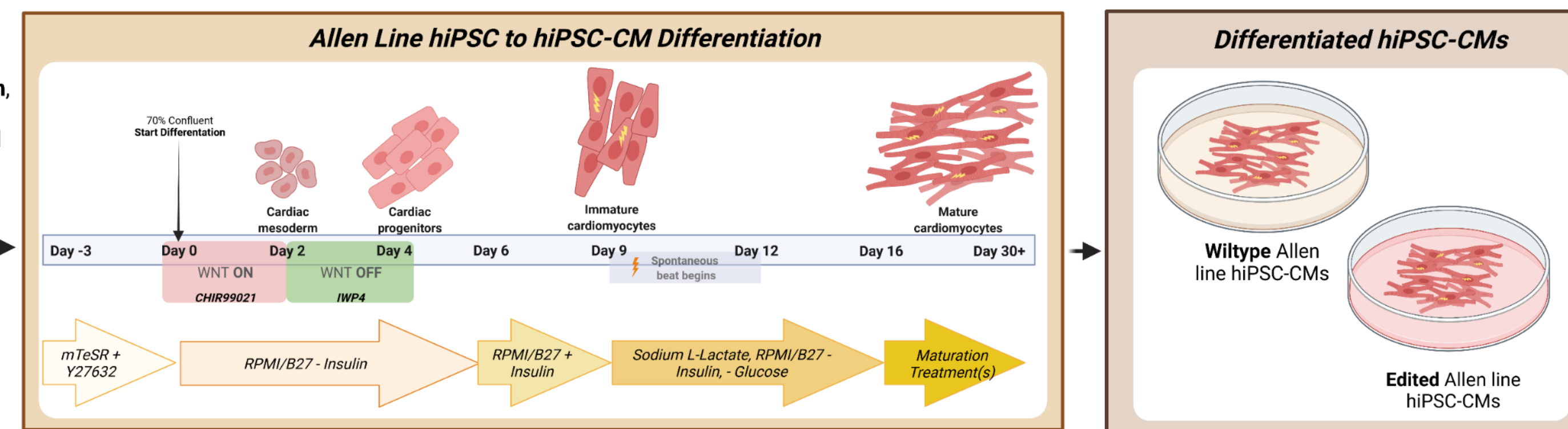
Develop a robust prime editing protocol in hiPSCs to study pro-arrhythmogenic gene variants in differentiated hiPSC-CMs

A high-fidelity and efficient workflow for developing variant hiPSC-CMs using PE3b-based Prime

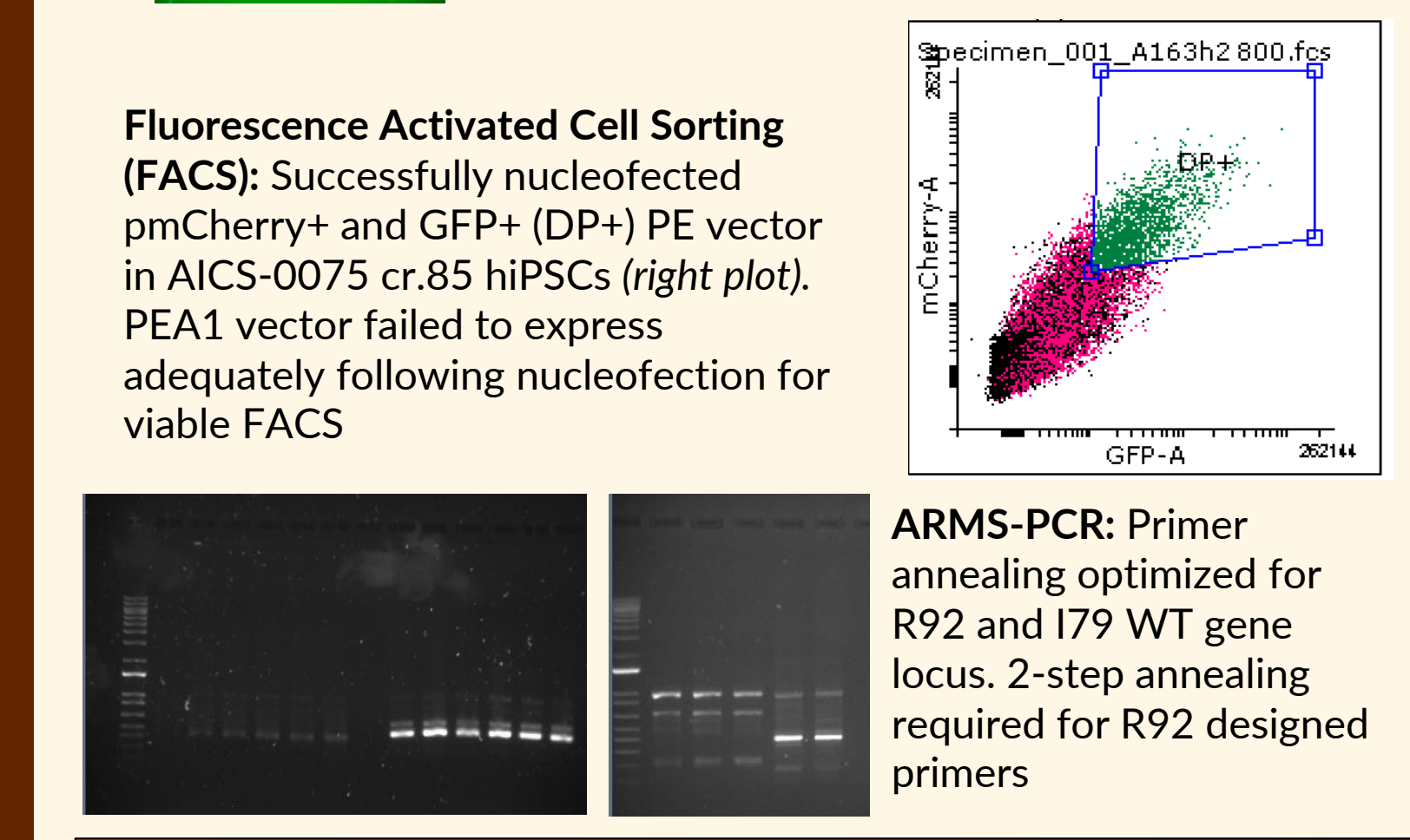
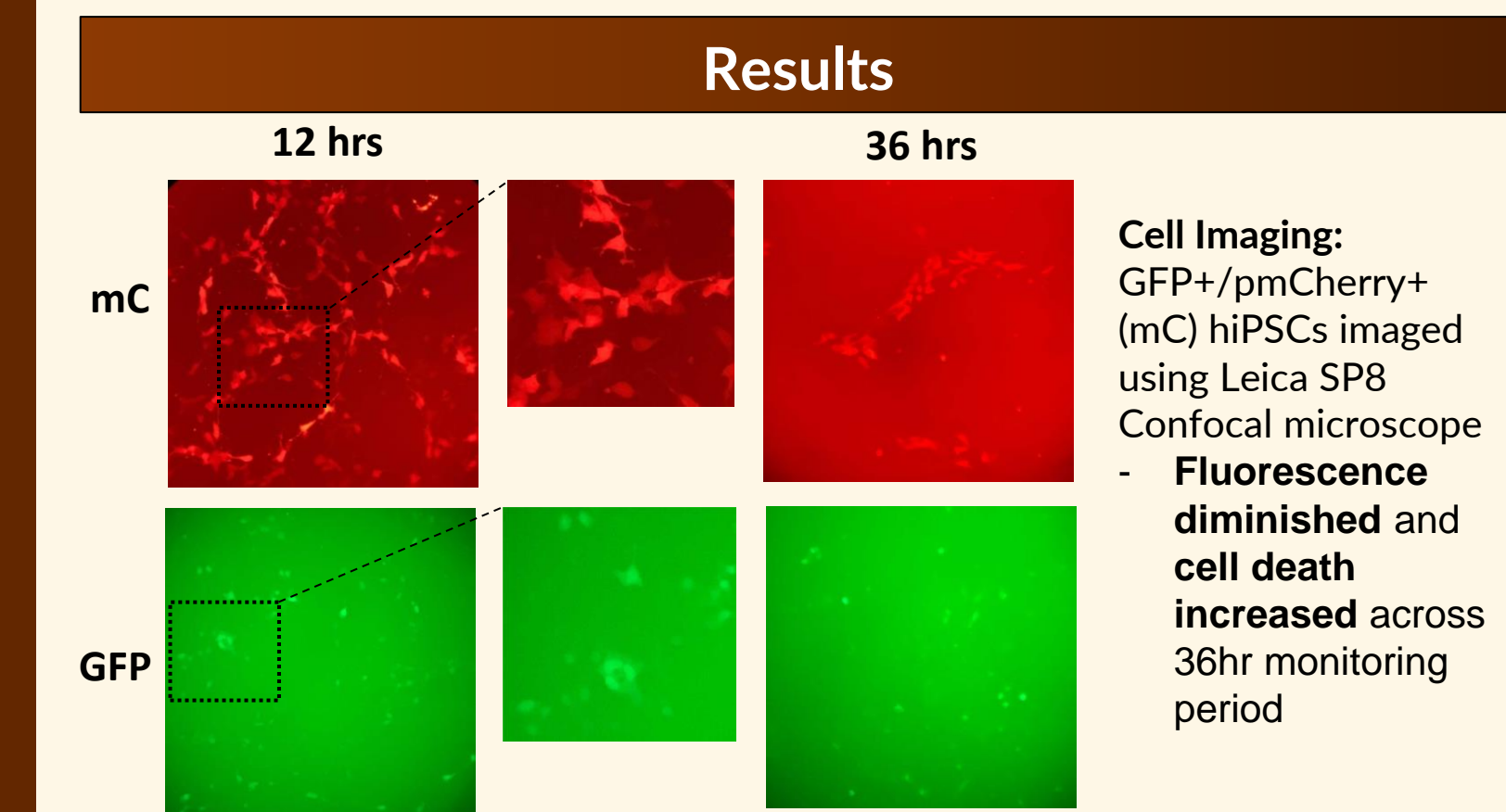
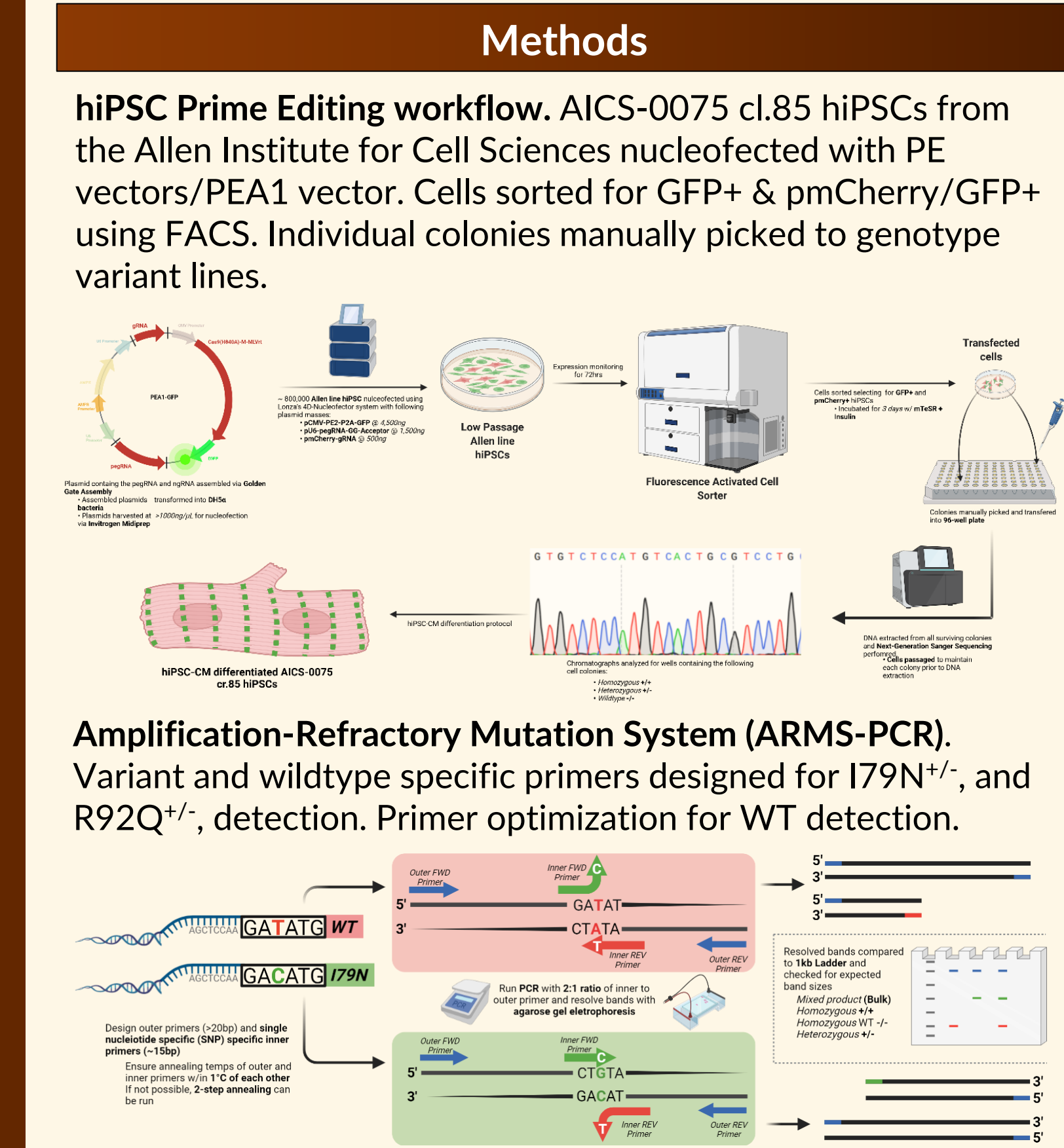
Editing



4 hiPSC line specific Differentiation, Lactate Selection, and subsequent Maturation Protocol(s)



5 Measurement of parameter variation due to introduced variant vs wildtype



- ## Summary
- Preliminary work established effective PE vectors & PEA1 nucleofection-based transfection and FACS based cell sorting for identifying and individualizing potentially edited hiPSCs
 - Waning expression and lack of effective PE expression due to rapid promoter silencing remain obstacles
 - ARMS-PCR optimized for WT hiPSCs at 179 and R92 *TNNT2* gene locus
 - High degree of variant specific primer optimization trial questions usability in research-based applications

Diagrams made using **bio RENDER**

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Acknowledgments



References

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 2 Chemello F, Chai AC, Li H, et al. Precise correction of Duchenne muscular dystrophy exon deletion mutations by base and prime editing. *SciAdv*. 2021;7(18):eabg4910. Published 2021 Apr 30. doi:10.1126/sciadv.abg4910