Shieldin Genetics SHLD1 CRISPR-Cas9 K/O



BACKGROUND:

DNA double strand breaks (DSBs) are a common type of DNA damage in both healthy and malignant/cancerous cells. Shieldin is a recently discovered four subunit complex involved in the DNA repair of DSBs, through a process called NHEJ. The gene interaction type between shieldin subunits was the initial proposed question, with the first step undertaken here; CRISPR-Cas9 editing of shieldin subunits.

METHODS



DISCUSSION

• CRISPR-Cas9 technology allowed the generation of cell lines with identical genomic background and SHLD1 knocked out. Future work could entail k/o of the second SHLD1 allele, SHLD2 alleles, and/or SHLD3 and REV7. These will be suitable for gene interaction analysis of the shieldin complex.



TIDE PCR product agarose gel electrophoresis

The figure to the right shows two bars at -1 and -10, indicating 1 and 10 base pair deletions. Both of these deletions are, hence, frameshift mutations. In this particular case, the k/o is homozygous, as suggested by the absence of a bar at indel size 0.



SHLD1 CRISPR K/O in Cas9+ Cells



The discordance plot details the level of alignment per base between the wild type (control) and the edited sample in the inference window (the region around the cute site), i.e. it shows the average amount of signal that disagrees with the reference sequence derived from the control trace file. On the plot, the green line and orange line should be close together before the cut site, with a typical CRISPR edit resulting in a jump in the discordance near the cutsite and continuing to remain far apart after the cut site (representing a high level of sequence discordance).



The Indel plot displays the inferred distribution of indels in the entire edited population of genomes. Hovering over each bar of the Indel plot shows the size of the insertion or deletion (+ or - 1 or more nucleotides), along with the percentage of genomes that contain it. Note: Each indel size represented in the Indel plot may not necessarily occur in the same sequence. The percentages of different indel sizes in the cell population are not the same as ICE/KO-Score scores.

•	SE	SEQUENCE																																																		
7%	G	A G	C	A	G	T	G	ст	т	T	G	G /		: c	Т	G	C	С	A 1	C	A	ł	-	-		-	-	-		-	Т	A	A	G 🖊	G	A	T	r a	C	G	г	c c	T	G		•	G	G	A	C C	C	A
6%	G	A (C C	A	G	T	G	ст	т	т	G	G /	۱ C	: C	Т	G	C	С	A 1	C	A	ł	-	C	G T	G	Т	G	A (A	Т	A	A	G 🖊	G	A	T	r a	C	G	Г	c c	т	G	C /	•	6 G	G	A	C C	C /	A

The contributions show the inferred sequences present in your edited population and their relative proportions (in contrast to the Indel plot (Indel Distribution tab) that does not specify sequence contributions). Cut sites are represented by black vertical dotted lines, and the wild-type

G C C A A G A A G C C A A G C C A A G A A G C C A A

EDITED SAMPLE 59 TO 124 E T G T C A G A G G A G C A G T G C T T T G G A C C T G C C A T C A¦C A T G A G A CT A C A T C C A G T A N G G C C C G C A C G G A G C A G T G C T T T G G A C C T G C C A T C A¦G C G T G T G A C A T A A G A G A T T A C G T C C T G C A G G s is the Sanger sequence view showing edited and wild-type (control) sequences in the region around the guide sequence. This show uence base calls from both the control and the experimental sample .ab1 files, which will contain mixed base calls. The horizontal bla inderlined region represents the guide sequence. The horizontal red underline is the PAM site. The vertical black dotted line represents the

The black bar indicates location of sgRNA cut site. Following the cut site, there is variation between nucleotide sequence when control (top) and edited (bottom) are compared.

REFERENCES:

actual cut site. Cutting and error-prone repair usually results in mixed sequencing bases after the cut.

Dev, H., Chiang, et al. (2018). Shieldin complex promotes DNA end-joining and counters homologous recombination in BRCA1-null cells. Nat Cell Biol, 20, 954– 965. <u>https://doi.org/10.1038/s41556-018-</u> 0140-1

Ghezraoui, H., et al. (2018). 53BP1 cooperation with the REV7-shieldin complex underpins DNA structure-specific NHEJ. Nature, 560, 122–127. https://doi.org/10.1038/s41586-018-0362-

Gupta, R., et al. (2018). DNA Repair Network Analysis Reveals Shieldin as a Key Regulator of NHEJ and PARP Inhibitor Sensitivity. Cell, 173(4), 972-988. <u>https://doi.org/10.1016/j.cell.2018.03</u> .050

Mirman, Z., et al. 53BP1–RIF1–shieldin counteracts DSB resection through CSTand Pol α -dependent fill-in. Nature, 560, 112-116. (2018). https://doi.org/10.1038/s41586-018-0324-

Noordermeer, S. M., et al. (2018). The shieldin complex mediates 53BP1dependent DNA repair. Nature, 560(7716), 117-121. <u>https://doi.org/10.1038/s41586-</u> <u>018-0340-7</u>te

@Setiaputra Lab, MBB Zahra Salimi @zsa74@sfu.ca

